

Determination of some non –steroidal anti-inflammatory drugs through quenching the fluorescence of lanthanide Tris complex

Mohamed. Rizk ^a, Safaa S. Toubar ^b, Mona S. Elshahed ^{a*}

^a Department of Analytical Chemistry, Faculty of Pharmacy, University of Helwan, Egypt

^b Faculty of Clinical Pharmacy, King Faisal University, Al-Hasa, KSA.

Received: 25/01/2010; Accepted: 10/05/2011

Abstract

A new, simple and sensitive spectrofluorimetric method is developed for the determination of Piroxicam (PX), Tenoxicam (TX) and Lornoxicam (LX). The proposed method involves the formation of complex with terbium (Tb³⁺) in the presence of Tris buffer. The quenching of terbium fluorescence due to complex formation is monitored at λ_{em} 540 nm after using λ_{ex} 237 nm. A linear relationship is found between the concentration of the studied drugs and the quenching of the fluorescence of terbium-Tris complex. The optimum reaction conditions are studied and optimized. The proposed method is applicable over the concentration range $1.509 \times 10^{-6} - 1.509 \times 10^{-5}$ M ($0.5-5 \mu\text{g mL}^{-1}$), $2.96 \times 10^{-6} - 1.48 \times 10^{-5}$ M ($1-5 \mu\text{g mL}^{-1}$) and $1.34 \times 10^{-6} - 1.34 \times 10^{-5}$ M ($0.5-5 \mu\text{g mL}^{-1}$) with mean percentage recovery 99.66 ± 3.3 , 100.79 ± 4.9 and 99.7 ± 1.1 for PX, TX and LX respectively. The Correlation Coefficient is 0.9983 (n=8), 0.9989 (n=5) and 0.9999 (n=6) with limit of detection 0.2, 0.25 and $0.2 \mu\text{g mL}^{-1}$ for PX, TX and LX respectively. The proposed method is applied successfully for the determination of the three drugs in their pharmaceutical formulations. The results are compared favorably with those of official or reference methods.

Keywords:

Piroxicam, Tenoxicam, Lornoxicam, Terbium and Spectrofluorimetry

1. Introduction

Piroxicam (4-hydroxy-2-methyl-N-2-pyridinyl-2H-1,2- benzothiazine-3-carboxamide 1,1- dioxide), Tenoxicam (4-hydroxy-2-methyl-N-2-pyridinyl-2H-tieno [2,3-e]-1,2-thiazine-3-carboxamide 1,1-dioxide) and Lornoxicam (6-Chloro- 4-hydroxy-2-methyl-N-2-pyridinyl-2H-tieno [2,3-e]-1,2-thiazine-3-carboxamide 1,1-dioxide), are non-steroidal anti-inflammatory and analgesic agents [1]. They belong to the class of oxicams; they are indicated in rheumatoid arthritis, osteoarthritis, extrarticular inflammation and acute gout [2]. Different methods were described for the determination of PX and or TX in pharmaceutical formulations and biological fluids. These methods include electro analytical methods such as voltammetry [3-6] and potentiometry [7, 8], spectrophotometric methods [9-12] and spectrofluorimetric methods [13-15]. The reported methods for determination of PX and or TX involve thin layer chromatography [16], high performance liquid chromatography [17-21] and electrophoresis [22]. Few methods were reported for determination of LX by spectrophotometry [23] and high- performance liquid chromatography [24-26].

* Corresponding Author

E-mail: mona_elshahed@yahoo.com; monaelshahed@gmail.com

ISSN: 1306-3057

Lanthanide ions (especially Tb^{3+} and Eu^{3+}) tend to form stable chelates with organic ligands. These lanthanide chelates are characterized by large Stokes shift, narrow emission bands and long fluorescence life times. The reaction of lanthanide ions with some organic compounds was used to improve the fluorimetric detection of certain organic analytes. This method is known as lanthanide –sensitized luminescence [27] and it was used for fluorimetric determination of some fluoroquinolones [28-30], tetracycline [31], anthranilic acid derivatives [32], ciprofloxacin [33] and DNA [34]. Some organic compounds can quench the background luminescence of these ions. The quenching effect is more important with the chloride than the nitrate salts of the lanthanides, because the probability of collisions leading to energy transfer is larger for the chloride salts [27]. This quenching phenomenon was reported for the fluorimetric determination of some cephalosporins [35]. This work utilizes the quenching effect of the studied compounds on the fluorescence of terbium (Tb^{3+}) in Tris buffer pH 8.5 for the determination of these compounds in bulk as well as in some of their pharmaceutical formulations.

2. Experimental

2.1. Apparatus

BIO-TEK Model SFM 25 single beam spectrofluorometer equipped with double monochromator system with reference beam compensation of source changes using 1cm quartz cell. Sensitivity adjusted to be 700 mV. Digital pH meter (Consort P-300) was used for adjustment of pH

2.2. Materials and Method

2.2.1. Reference samples

PX, TX and LX pure samples, kindly supplied by (EL-Obour Modern Pharmaceutical Industries Co, Egypt). The % purity of PX and TX was checked according to B.P[36] and was found to be (99.7) and (99.1) respectively. The purity of LX was provided by the company to be 99.6% and used as supplied.

2.2.2. Market samples

Felden tablet (Pfizer Co., Egypt) each tablet contains 20 mg of PX. Feldoral capsule, Feldoral suppository and Feldoral ampoule (Sedico Co., Egypt) are labeled to contain 20 mg of PX per capsule, suppository or ampoule. Tenocam capsule (Kahira Co., Egypt) each capsule contains 20 mg of TX. Xefo tablet (Manufactured by October Pharma S.A.E.6 October city for Egyptian Promotor center for Pharmaceuticals S.A.E. under license of Nycomed, Austria) each tablet contains 8 mg of LX.

2.2.3. Reagents

All chemicals used are of analytical grade and are used without further purification. Terbium (III) Chloride hexahydrate 99.9% (Aldrich, Germany), 2×10^{-3} M methanolic solution was prepared. Tris buffer [Tris (hydroxyl methyl) amino methane] was obtained from (Nice Chemicals, India), 0.1 mol L⁻¹ methanolic solution was prepared [36]. Methanol HPLC grade 99.9 % was obtained from (Sigma, Germany). Sodium hydroxide pellets was obtained from (Winlab, Leicestershire, U.K.). Sodium chloride was obtained from (Winlab, Leicestershire, U.K.). Hydrochloric acid HPLC grade 37 % was obtained from (Rediel –De-Haen, Germany).

2.3. Preparation of Sample Solutions

PX, TX and LX stock sample solutions (0.8 mg mL⁻¹) were prepared by dissolving 20 mg of each drug in 2 ml 0.2 mol L⁻¹ NaOH in 25 mL volumetric flask and completed to volume with methanol. A working sample solution (40 µg mL⁻¹) of each drug was freshly

prepared by appropriate dilution of the stock solution with methanol. The stock solutions were kept in a refrigerator at approximately 4°C and remained stable for at least 1 month [14].

2.4. Construction of Calibration Graphs

1.0 mL of Tris buffer pH 8.5 ± 0.2 was transferred in a series of 10 mL volumetric flasks followed by 1.0 mL of terbium solution (2×10^{-3} M). Aliquots of the studied drugs working solutions equivalent to 5-50 µg for PX and LX and 20-50 µg for TX were added. The solutions were mixed well and allowed to stand for 15 minutes then they were completed to volume with methanol. The relative fluorescence intensity was measured at λ_{540} nm after excitation at λ_{237} nm. The final concentration of the studied drugs was plotted versus the calculated fluorescence quenching (F^0/F) to obtain the standard calibration graphs. Alternatively the corresponding regression equations (Stern- Volmer equation) [37] were derived.

2.5. Note

F^0 : fluorescence of 0.2 m M solution of Tb^{3+} in Tris buffer and methanol without drug
 F : fluorescence of 0.2 m M solution of Tb^{3+} in Tris buffer and methanol after addition of the drug

2.6. Analysis of Dosage Forms

2.6.1. Felden and Xefo tablet:

Ten tablets were weighed and reduced to a fine powder .An amount equivalent to 20 mg of either PX or LX was accurately weighed and transferred to 25 mL volumetric flask, dissolved in 2 mL 0.2 mol L⁻¹ NaOH and made up to volume with methanol. The solution was sonicated for 15 minutes, filtered with rejection of the first few milliliters. The produced solution (0.8 mg mL⁻¹) of the studied drug was used to proceed as under preparation of sample solution.

2.6.2. Feldoral and Tenocam capsules:

The contents of not less than ten capsules were transferred to a suitable tared container and mixed well. An accurately weighed amount equivalent to 20 mg of PX or TX was transferred to 25 mL volumetric flask, dissolved in 2 mL 0.2 mol L⁻¹ NaOH and completed to volume with methanol. The solution was sonicated for 15 minutes, filtered with rejection of the first few milliliters .The produced solution is claimed to contain 0.8 mg mL⁻¹. The steps described under preparation of sample solution were followed.

2.6.3. Feldoral suppositories

Ten suppositories were weighed, cut into small pieces and transferred to a small porcelain dish. They were melted by stirring in a water bath to homogenize and then cooled. A portion equivalent to 20 mg PX was weighed into a beaker, melted and dissolved in 2 mL 0.2 mol L⁻¹ NaOH and methanol by stirring at 60 °C for 5 minutes. The solution was cooled, filtered and diluted to 25 mL with methanol to produce stock solution containing 0.8 mg mL⁻¹ PX. The produced solution was used to proceed as under preparation of sample solution.

2.6.4. Feldoral ampoules

The contents of ten ampoules were mixed and a volume equivalent to 20 mg PX was transferred to 25 mL volumetric flask, mixed with 2 mL 0.2 mol L⁻¹ NaOH and completed to volume with methanol to obtain stock solution containing 0.8 mg mL⁻¹ PX. The steps described under preparation of sample solution were followed.

Suitable aliquots of the above prepared solutions of different pharmaceutical preparations were treated as described under construction of calibration graphs. The results were calculated for each preparation from the corresponding regression equation.

3. Results and Discussion

3.1. Fluorescence spectral characteristics

The solution of terbium chloride in Tris buffer and methanol shows intense fluorescence, when excited at λ_{237} nm, two emissions bands at λ_{484} and λ_{540} nm are observed as shown in Fig.1. These bands are characteristic of terbium due to $^5D_4-^7F_6$ and 7F_5 transitions respectively [38]. The fluorescence spectra of TX [$5\mu\text{g mL}^{-1}$] in methanol are represented in Fig.2. Weak signals are obtained at λ_{469} , λ_{651} , λ_{667} nm after excitation at λ_{237} , λ_{327} , λ_{337} nm respectively. Fig.1 and 2 show that, there is no interference from the fluorescence of the studied drugs on the emission peak of Tb^{3+} -Tris complex at λ_{540} nm, so it is the wavelength chosen for fluorescence measurement. At λ_{484} nm the fluorescence of the studied drugs interferes with that of Tb^{3+} -Tris complex, resulting in non-quantitative quenching effect. Reviewing the literature revealed that the weak native fluorescence of the studied drugs has no useful analytical application [14], Therefore the complexation reaction between terbium and the studied drugs is carried out to improve the fluorimetric determination of these drugs. Quantitative measurements are made by excitation at λ_{237} nm and measuring the fluorescence at λ_{540} nm where the studied drugs show no interference. When the studied drugs are added to Tb^{3+} they react with terbium forming stable complexes with no fluorescent properties which cause quenching of the fluorescence intensity of methanolic Tb^{3+} -Tris complex. This quenching is proportional to the concentration of the added drug. The quenching of the fluorescence signal of Tb^{3+} in Tris buffer after complexation with LX and PX are shown in Fig. 3 and 4, respectively.

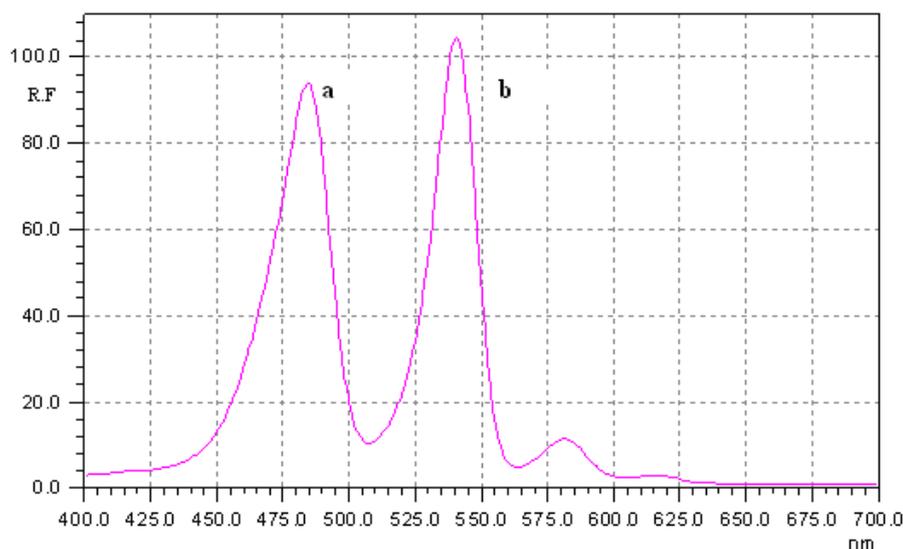


Fig 1. The emission spectra of terbium-Tris complex at [λ_{484} (a) and λ_{540} (b) after excitation at λ_{237}]

N.B. concentration of TbCl_3 is (0.2 mM)

R.F. = the relative fluorescence

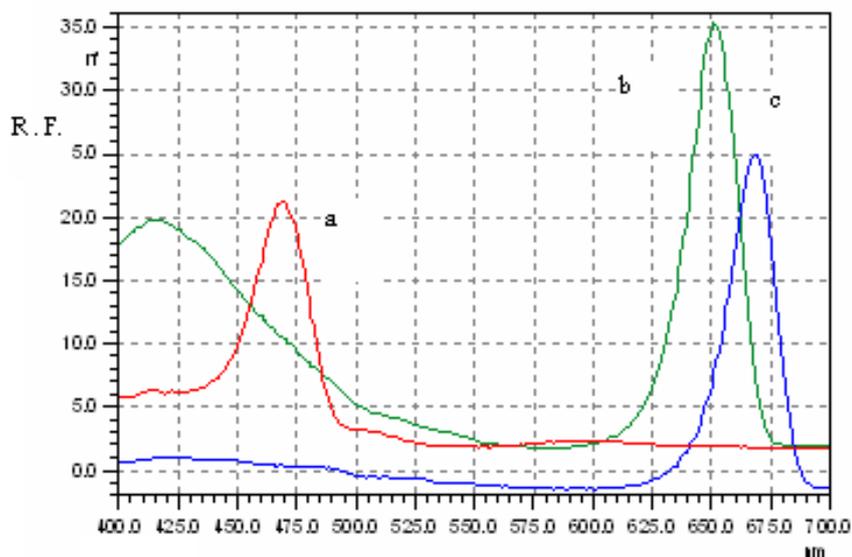


Fig 2. The emission spectra of TX ($5\mu\text{g mL}^{-1}$) in methanol at (a) $\lambda=469$, (b) $\lambda=651$ and (c) $\lambda=667$ nm after excitation at $\lambda=237$, $\lambda=327$ and $\lambda=335$, respectively

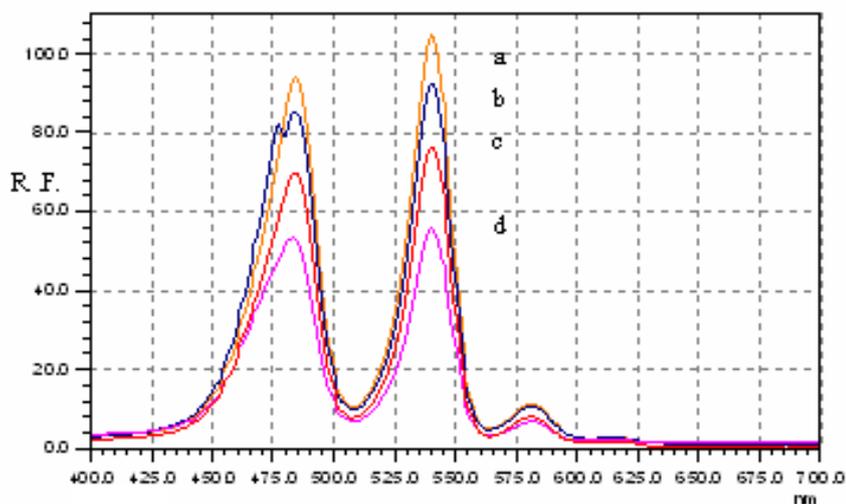


Fig 3. Stern Volmer plot of 1-(0.2mM TbCl_3 in tris buffer and methanol (a)) 2-(0.2mM TbCl_3 in tris buffer and methanol + [0.5 (b), 2.0 (c) and 5.0 (d) $\mu\text{g mL}^{-1}$ of LX])

N.B. concentration of TbCl_3 is (0.2mM)

R.F. = the relative fluorescence

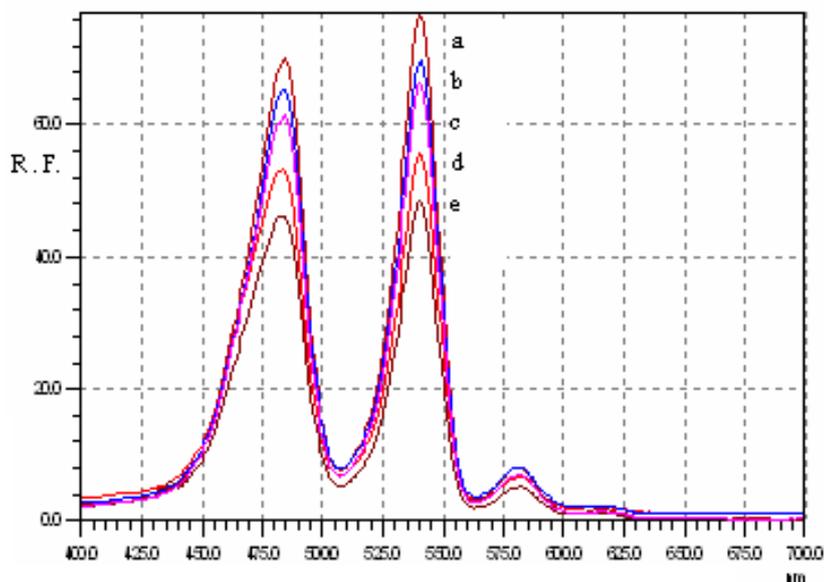


Fig 4. Stern Volmer plot of 1- {0.2mM TbCl₃ in tris buffer and methanol (a) 2- (0.2mM TbCl₃ in tris buffer and methanol + [1.0 (b), 1.5 (c), 3.0 (d) and 5.0 (e) μg mL⁻¹ of PX])

N.B. concentration of TbCl₃ is (0.2mM)

R.F. = the relative fluorescence

3.2. Optimization of the experimental conditions

Different experimental parameters affecting the relative fluorescence of Tb³⁺-Tris complex and complex formation between the studied compounds and terbium were carefully studied and optimized. Such factors were changed individually, while others were kept constant. These factors include Volume of terbium chloride solution, Effect of pH and Effect of time on complex formation.

3.2.1. Volume of terbium chloride solution

The fluorescence of Tb³⁺ in Tris buffer and methanol at λ_{em} 540 nm (λ_{ex} 237 nm) was measured with increasing concentration of terbium and was found to increase quantitatively with terbium concentration as shown in Fig. (5). It was found that 1 mL of (2 × 10⁻³ mol L⁻¹) of Tb³⁺ is appropriate for reasonable fluorescence intensity.

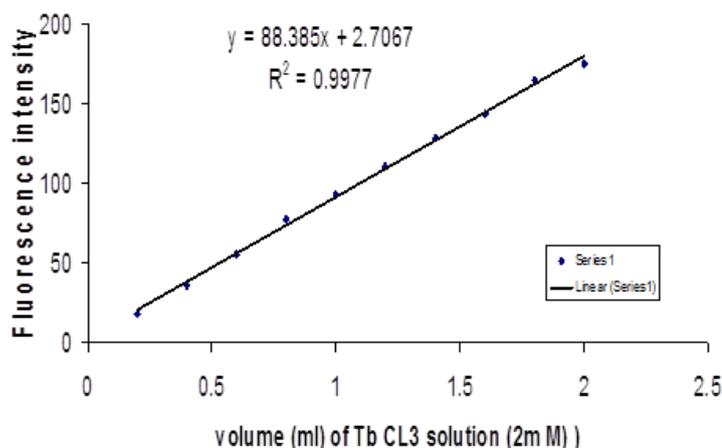


Fig 5. Calibration graph of the fluorescence of terbium chloride-Tris complex in methanol

3.2.2. Effect of pH

The influence of pH on relative fluorescence of Tb^{3+} and complex formation between terbium and the studied compounds was studied using different types of buffers covering wide range of pH. The results are represented in Fig. (6) and can be summarized as follow.

- *Acetate buffer*: the reaction was studied in acetate buffers of pH 4.7 and 5.5 where the fluorescence of terbium before and after addition of the drug was nil.
- *Borate buffer*: Solution of Tb^{3+} in borate buffer at different pH values (7.0, 8.0, 9.0 and 10.0) ± 0.2 showed intense fluorescence at λ_{540} nm. Quenching of this fluorescence upon addition of the studied drug was observed and was found to increase with increasing drug concentration but slight turbidity was observed which may be attributed to formation of terbium (III) hydroxide and caused measurements to be none reproducible.
- *Tris buffer*: The quenching of terbium fluorescence after addition of TX ($3 \mu\text{g mL}^{-1}$ final concentration) was measured in different solutions of Tris buffer pH (8.0, 9.0, and 10.0) ± 0.2 . The study revealed no significant difference in fluorescence quenching with change of pH. Tris buffer of pH 8.5 ± 0.2 was used for calibration.

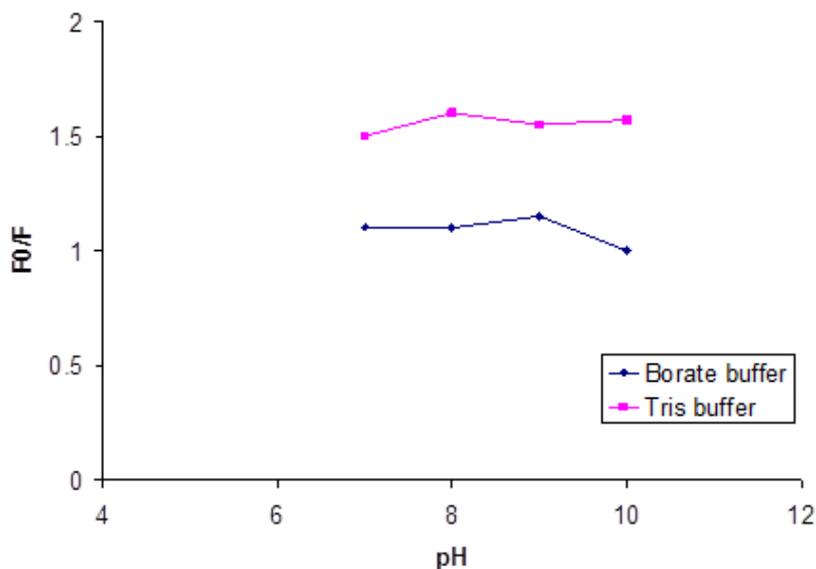


Fig 6. Effect of pH on complex formation between Terbium and the studied drugs (TX $3 \mu\text{g mL}^{-1}$ final concentration)

3.2.3. Effect of time on complex formation

The fluorescence of terbium–Tris solution after addition of TX ($2 \mu\text{g mL}^{-1}$ final Concentration) was monitored after several time intervals. It was found that the complex was immediately formed and remained stable for at least 2 hours as shown in Fig 7. The fluorescence all over this study was measured after 15 minutes at room temperature.

3.2.4. Analytical features

After optimizing the conditions, the calibration graphs of PX, TX and LX were constructed by plotting the calculated fluorescence quenching of terbium solution (F^0/F) versus final concentration in $\mu\text{g mL}^{-1}$. The fluorescence–concentration plots were linear over the concentration range $1.509 \times 10^{-6} - 1.509 \times 10^{-5}$ M ($0.5-5 \mu\text{g mL}^{-1}$), $2.96 \times 10^{-6} - 1.48 \times 10^{-5}$ M ($1-5 \mu\text{g mL}^{-1}$) and $1.34 \times 10^{-6} - 1.34 \times 10^{-5}$ M ($0.5-5 \mu\text{g mL}^{-1}$) with mean percentage recoveries

99.66 ± 3.4 (n=8), 100.79 ± 4.9 (n=5) and 99.70 ± 1.1 (n=6) for PX, TX and LX respectively. Analysis of the data gave the following regression equations:

$$F^0/F = 0.1285 C + 1.0128 \quad (r = 0.9983) \quad \text{for PX}$$

$$F^0/F = 0.2414 C + 0.902 \quad (r = 0.9989) \quad \text{for TX}$$

$$F^0/F = 0.147 C + 1.024 \quad (r = 0.9999) \quad \text{for LX}$$

Where F^0 is the fluorescence of 0.2 m M solution of Tb^{3+} in Tris buffer and methanol without drug, F is the fluorescence of 0.2 m M solution of Tb^{3+} in Tris buffer and methanol after addition of the drug, C is the concentration of the studied drug in ($\mu\text{g mL}^{-1}$) and (r) is the correlation coefficient.

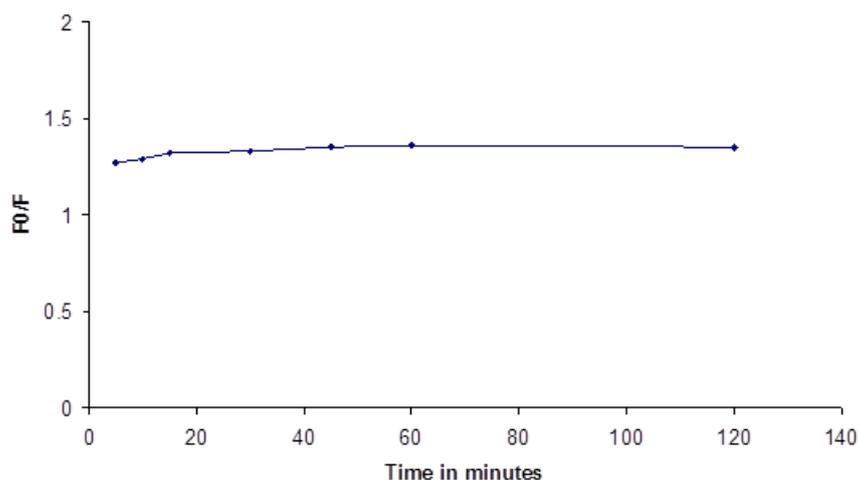


Fig 7. Effect of time on complex formation between Terbium and the studied drugs (TX $2 \mu\text{g mL}^{-1}$ final concentration) in Tris buffer

The limit of quantification (LOQ) was determined by establishing the lowest concentration that can be measured according to ICH Q2(R1) recommendations [39], below which the calibration graph is non linear (based on visual evaluation), while the limit of detection (LOD) was determined by evaluating the lowest concentration of the analyte that can be readily detected. The results of LOD and LOQ of the studied drugs by the proposed method are abridged in Table 1.

The proposed method was evaluated by studying the accuracy as percent relative error and precision as percent relative standard deviation. The results are abridged in Table 1. Statistical analysis [41] of the results obtained by the proposed method and the official method for PX [40] and the reference methods for TX [17] and LX [25] using Student's t-test and variance ratio F-test, shows no significant difference between the performance of our work and these methods regarding the accuracy and precision, respectively (Table 2).

3.3. Validation of the method

3.3.1. Linearity

The proposed method was tested for linearity, specificity, accuracy and precision. Linear regression equations were obtained. The regression plots showed linear dependence of fluorescence quenching of terbium solution on concentration of the studied drugs over the range cited in Table 1. The small values of the %RSD and %Er point out to the low scattering of the points around the calibration curve and high accuracy and precision of the proposed method.

Table 1. Performance data for the studied NSAID drugs by the proposed fluorescence quenching method.

Parameter	NSAID drugs		
	PX	TX	LX
Working range	0.5–5.0 $\mu\text{g mL}^{-1}$	1.0–5.0 $\mu\text{g mL}^{-1}$	0.5–5.0 $\mu\text{g mL}^{-1}$
Mean % Recovery	99.66	100.79	99.70
\pm S. D.	3.30	4.93	1.10
Relative standard deviation (%RSD)	3.30	4.90	1.10
Regression equation	$F^0/F = 0.1285 C + 1.0128$	$F^0/F = 0.2414 C + 0.902$	$F^0/F = 0.147 C + 1.024$
Correlation coefficient (r)	0.9983	0.9989	0.9999
Standard deviation of the residuals ($S_{y/x}$)	0.0123	0.0201	0.0029
Percentage error %Er	1.17	2.18	0.45
Limit of Quantitation (LOQ)	0.5 $\mu\text{g mL}^{-1}$	1.0 $\mu\text{g mL}^{-1}$	0.5 $\mu\text{g mL}^{-1}$
Limit of Detection (LOD)	0.2 $\mu\text{g mL}^{-1}$	0.25 $\mu\text{g mL}^{-1}$	0.2 $\mu\text{g mL}^{-1}$

F^0 : fluorescence of 0.2 m M solution of Tb^{3+} in Tris buffer and methanol without drug

F: fluorescence of 0.2 m M solution of Tb^{3+} in Tris buffer and methanol after addition of the drug

C = Concentration in $\mu\text{g mL}^{-1}$

LOQ and LOD are measured Based on Visual Evaluation

Table 2. Statistical analysis of the results obtained for the studied drugs in pure form by the proposed spectrofluorimetric method and comparison methods

Studied drug	Proposed method		comparison method	
	concentration taken ($\mu\text{g mL}^{-1}$)	concentration found ($\mu\text{g mL}^{-1}$)	Recovery	% Recovery
PX	0.5	0.499	99.2	99.00
	1.0	0.995	99.5	98.00
	1.5	1.410	94.00	97.50
	2.0	1.970	98.50	95.30
	2.5	2.620	104.80	101.20
	3.0	2.990	99.67	103.60
	4.0	4.130	103.25	
	5.0	4.880	97.60	
Mean (X) \pm S. D.	99.66 \pm 3.32		99.1 \pm 2.9	
Variance	11.02		8.41	
Variance ratio	1.31 (4.88) **			
F-test				
Student's t-test	0.33 (1.78) **			
TX	1.0	1.09	108.90	95.60
	2.0	1.92	96.10	100.90
	3.0	2.93	99.76	103.20
	4.0	4.01	100.20	97.30
	5.0	5.05	101.00	102.00
Mean (X) \pm S. D.	100.79 \pm 4.93		99.8 \pm 3.2	
Variance	24.3		10.24	
Variance ratio (F-test)	2.37 (6.39) **			

Table 2. contiued

Studied drug	Proposed method			comparison method
Student's t-test	0.38 (1.85)**			
LX	0.5	0.49	98.00	97.50
	1.0	0.99	99.00	98.50
	2.0	2.01	100.5	96.60
	3.0	3.03	101.00	96.90
	4.0	3.98	99.5	101.00
	5.0	5.01	100.2	
_Mean(X) ± S. D.	99.7 ± 1.1		98.1 ± 1.78	
Variance	1.21		3.17	
Variance ratio (F-test)	2.62 (5.19) **			
Student's t-test	1.82 (1.83) **			

** The value in brackets are the tabulated ones at P =0.05

3.3.2. Accuracy and precision

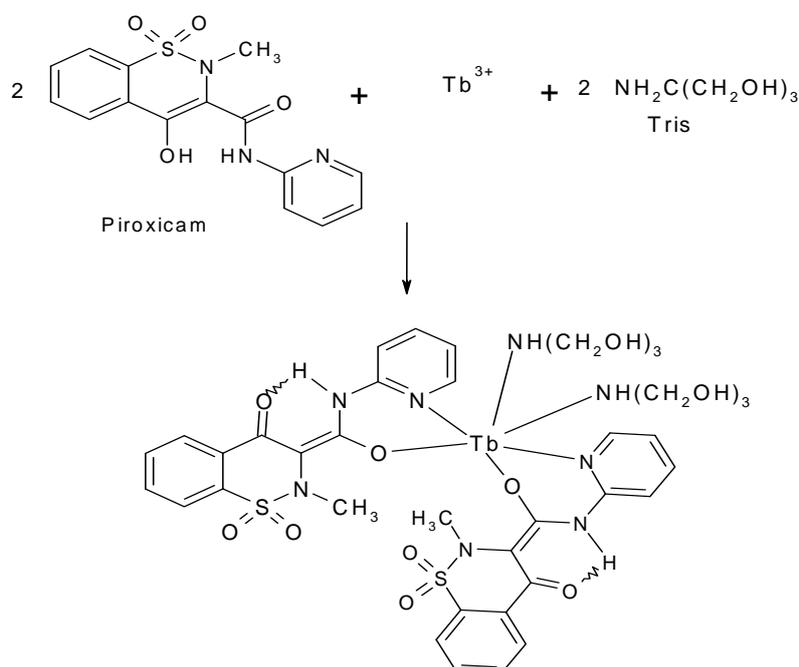
The results of the inter-day and intra-day accuracy and precision of the method have been summarized in Table 3. The inter-day and intra-day precisions were evaluated through replicate analysis of the studied drugs in pure form using different concentrations (2.0, 3.0, 4.0 $\mu\text{g mL}^{-1}$) and each concentration was measured three times a day for three consecutive days. The precision of the proposed method was fairly high, as indicated by the low values of SD and %RSD, respectively. Also the inter-day and intra-day accuracy was proved by the low values of %Er.

Table 3. Precision data for the studied drugs by the proposed fluorescence quenching method.

Parameter	Intra-day precision (Repeatability)			Inter-day precision (intermediate precision)		
	2.0	3.0	4.0	2.0	3.0	4.0
concentration taken ($\mu\text{g mL}^{-1}$)	2.0	3.0	4.0	2.0	3.0	4.0
Recovery of PX, %	100.40	101.80	99.3	98.50	99.80	102.80
	97.10	99.80	97.1	102.70	105.30	103.40
	96.70	105.10	98.4	95.50	101.90	100.40
Mean (\bar{X})	98.10	102.20	98.40	98.90	102.30	102.20
± SD	2.10	2.70	1.80	3.60	2.80	1.60
%RSD	2.10	2.60	1.80	3.70	2.70	1.60
%Er	1.20	1.50	1.10	2.10	1.60	0.90
%Recovery of TX	96.50	97.50	102.4	94.1	97.40	100.90
	97.20	96.90	105.2	97.60	100.30	95.20
	94.80	99.60	100.00	101.20	94.50	103.90
Mean (\bar{X})	96.20	98.00	102.50	97.60	97.40	100.00
± SD	1.20	1.36	2.60	3.60	3.10	4.40
%RSD	1.30	1.41	2.50	3.70	3.20	4.40
%Er	0.70	0.80	1.50	2.10	1.80	2.50
Recovery of LX, %	103.50	100.30	99.3	103.6	100.7	99.50
	99.10	98.30	96.00	100.2	104.2	96.00
	96.70	102.90	100.10	101.3	102.3	95.20
Mean (\bar{X})	99.80	100.50	98.50	101.7	102.4	96.90
± SD	3.50	2.30	2.20	1.70	1.80	2.30
%RSD	3.50	2.30	2.20	1.70	1.80	2.40
%Er	2.00	1.30	1.30	0.97	1.0	1.40

3.4. Proposed mechanism of the reaction

In lanthanide-sensitized luminescence the intense luminescence originates from an intra molecular energy transfer through the excited triplet state of the ligand to the emitting resonance level of the ion followed by radiative emission from the cation. The efficiency of the energy transfer depends on the matching between the triplet level of the organic compound and the resonance level of the ion. The energy of the triplet level should be close to but higher than, that of the resonance level of the ion. In some instances, when the organic compound has a triplet state level below the excited state level of the lanthanide ion the organic compound can quench the back ground luminescence of the ion [27]. Several studies were made on TX and PX as chelating agents with uranyl (UO₂ II), Fe III, Co II, Ni II, Cu II and other metals [42-44]. These studies mentioned that TX and PX act as neutral bidentate ligand coordinated to the metal ion via the pyridine -N and carbonyl-O groups. The structure of the complex formed between the studied drugs and Tb³⁺ is proposed to be as follow.



3.5. Application

3.5.1. Analysis of pharmaceutical formulations

The proposed method was applied for determination of PX in tablets, capsules, suppositories and ampoules, TX in capsules and LX in tablets. The results are shown in Tables 4 and 5. The results of the proposed and reference methods were compared in accordance with the Student's t-test and variance ratio F-test [41]. There were no significant differences between the calculated and tabulated values at P = 0.05, demonstrating that the proposed method is as accurate and precise as the respective official and reference methods.

4. Conclusion

The data of the results given by this proposed procedure are indicative for higher sensitivity and reasonable selectivity. Furthermore, these findings are favorably comparable to other methods.

Table 4. Statistical analysis of the results obtained for PX in some pharmaceutical preparations using the proposed fluorescence quenching method and official method

Parameter	Feldoral capsule		Felden tablet		Feldoral suppository		Feldoral ampoule	
	Proposed method	Official method	Proposed method	Official method	Proposed method	Official method	Proposed method	Official method
concentration taken μgmL^{-1}	2.0	40.0	2.0	40.0	2.0	40.0	2.0	40.0
Recovery of PX, %	95.70	94.0	100.30	106.3	105.10	102.60	105.90	103.30
	98.95	96.50	99.60	102.00	104.40	107.30	102.50	107.30
	100.30	95.30	96.30	100.3	102.20	104.95	103.90	107.00
Mean (X)	98.30	95.30	98.70	102.86	103.90	104.95	104.10	105.90
\pm S. D.	2.40	1.26	2.1	3.1	1.50	2.30	1.70	2.23
Variance	5.76	1.59	4.41	9.61	2.25	5.29	2.89	4.97
Variance ratio F-test	3.6		2.18		2.35		1.72	
Student's t-test	1.92		1.93		0.66		1.11	

N.B. Tabulated t and F values at P = 0.05 are t = 2.132 and F = 19.00

Table 5. Statistical analysis of the results obtained for TX and LX in some pharmaceutical preparations using the proposed fluorescence quenching method and comparison methods:

Parameter	Tenocam capsule (TX)		Xefo tablet (LX)	
	Proposed method	Comparison method [17]	Proposed method	Comparison method [25]
concentration taken μgmL^{-1}	3.0	7.50	3.0	1.0
Recovery, %	96.30	97.80	99.80	96.00
	101.90	95.30	98.50	97.50
	100.20	100.10	96.60	100.60
	99.40			
	97.95			
	101.10			
Mean (X)	99.47	97.70	98.30	98.03
\pm S. D.	2.00	2.40	1.60	2.35
Variance	4.00	5.76	2.56	5.29
Variance ration F-test	1.44	(5.79) **	2.10	(19.00) **
Student's t-test	1.09	(1.894) **	0.165	(2.132) **

N.B. ** The value in brackets are the tabulated ones at P = 0.05 N

References

- O'Neil M J (2006) The Merck Index , Fourteenth Edition , MERK & CO., INC. , Whitehouse station , NJ , USA , P 967 , 1294 , 1573
- Sweetman S C (2002) Martindale : the complete drug reference , Thirty-third Edition, Pharmaceutical Press , Great Britain , P 50 , 79, 87
- Acuna J A, De-La-Fuente C, Vazquez M D and Tascon M L (1993) Voltammetric determination of Piroxicam in micellar media by using conventional and surfactant chemically modified carbon paste electrodes. Talanta 40 (11) : 1637
- Gonzalez M, Vasquez M D, Tascon M L and Sanchez- Batanero P (1994) Contribution to the electrochemical study of Piroxicam in different aqueous-organic

- media and electrodes by using polarographic and voltammetric techniques. *Electroanalysis* (NY) 6: 497
5. Atokpar Z and Tuncel M (1996) The polarographic determination of Tenoxicam in pharmaceutical preparations. *Anal Lett* 29 (13): 2383
 6. Ozaltin N (2000) Differential pulse polarographic determination of Tenoxicam in pharmaceuticals and added to blood. *Anal Chem Acta* 406 (2): 183
 7. Khalil S, Borham N and El-Reis M A (2000) Piroxicam and Tenoxicam selective membrane sensors. *Anal Chem Acta* 414 (1-2): 215
 8. Mohamed H A, Wadood H M A and Farghaly O A (2002) Potentiometric and spectrofluorimetric studies on complexation of tenoxicam with some metal ions. *J Pharm Biomed Anal* 28 (5): 819
 9. Carcia M S, Sanchez-Pedreno C, Albero M I and Gimenez M J (1999) Flow-injection spectrophotometric method for determination of tenoxicam. *J Pharm Biomed Anal* 21(4): 731.
 10. El-Rries M A (1998) Spectrophotometric determination of piroxicam and tenoxicam in pharmaceutical preparations using uranyl acetate as chromogenic agent. *Anal Lett* 31(5): 793.
 11. Amin A S (2002) Spectrophotometric determination of piroxicam and tenoxicam in pharmaceutical formulations using alizarine. *J Pharm Biomed Anal* 29(4): 729
 12. Al-Momani I F (2006) Indirect flow –injection spectrophotometric determination of meloxicam, tenoxicam and piroxicam in pharmaceutical formulations. *Anal Sci* 22(12): 1611.
 13. Escandar G M, Bristol A J and Compiglia A D (2002) Spectrofluorimetric method for determination of piroxicam and pyridoxine. *Anal Chem Acta* 466 (2): 275
 14. Alkindy S M Z, Al-Wishahi A and Suliman F E O (2004) A sequential injection method for determination of piroxicam in pharmaceutical formulations using europium sensitized fluorescence. *Talanta*. 64 (5): 1343.
 15. Taha E A, Salama N N and Fattah Lel S (2006) Spectrofluorimetric and spectrophotometric stability indicating methods for determination of some oxicams using 7-chloro-4-nitrobenz-2-oxa-1,3-diazole(NBD-Cl). *Chem Pharm Bull (Tokyo)*, 54 (5): 653.
 16. Cercelius A, Clench M R, Richards D S and Parr V (2004) Quantitative determination of piroxicam by TLC-MALDI TOF MS. *J Pharm Biomed Anal* 35 (1): 31.
 17. Carlucci G, Mazzeo P and Palumbo G (1992) Determination of tenoxicam in human plasma using solid-phase extraction and high-performance liquid chromatography with ultra-violet detection. *J Liq Chromatogr* 15 (4): 683.
 18. Mason J L and Hobbs G J (1995) Simple method for the analysis of tenoxicam in human plasma using high-performance liquid chromatography. *J Chromatogr Biomed. Appl* 665 (2): 410.
 19. Joseph-Charles J and Bertucat M (1999) High-performance liquid chromatography analysis of non-steroidal anti-inflammatory oxicams in pharmaceutical preparations. *J Liq Chromatogr Relat Technol* 22 (3): 2009.

20. Abdel-Hamed M E (2000) LC-MS analysis of selected sulfur-containing non-steroidal anti-inflammatory agents: application to pharmaceutical products. *J Liq Chromatogr Relat Technol* 23 (20): 3095.
21. Ji H Y, Lee H W, Kim Y H, Jeon D W and Lee H S (2005) Simultaneous determination of piroxicam, meloxicam and tenoxicam in human plasma by liquid chromatography with tandem mass spectrometry. *J Chromatogr B* 826 (1-2): 214.
22. Donato M G, Baeyens W, Van-den-Bossche W and Sandra B (1994) Determination of non-steroidal anti-inflammatory drugs in pharmaceuticals by capillary zone electrophoresis and micellar electrokinetic capillary chromatography. *J Pharm Biomed Anal* 12 (1): 21.
23. Nemutlu E, Demircan S and Kir S (2005) Determination of lornoxicam in pharmaceutical preparations by zero and first order derivative UV spectrophotometric methods. *Pharmazie* 60 (6): 421.
24. Suwa T, Hurano H, Shinohara Y and Kokaysu J (1993) Simultaneous high-performance liquid chromatographic determination of lornoxicam and its 5'-hydroxyl metabolite in human plasma using electrochemical detection. *J Chromatogr Biomed Appl* 617 (1): 105.
25. Radhofer-Welte S and Dittrich P (1998) Determination of the novel non-steroidal anti-inflammatory drug lornoxicam and its main metabolite in plasma and synovial fluid. *J Chromatogr Biomed Appl* 707: 151.
26. Taha E A, Salama N N and Abdel Fattah Lel S (2004) Stability indicating chromatographic methods for the determination of some oxicams. *J AOAC Int* 87 (2): 366.
27. George J (1993) Lanthanide-sensitized luminescence and applications to the determination of organic analytes. *Analyst* 118: 1481.
28. Rizk M, Belal F, Aly F A and El-Enany N M (1997) Fluorimetric determination of nalidixic acid in formulations and biological fluids through ternary complex formation. *Anal Lett* 30 (10): 1897.
29. Ocana J A, Callejon M and Barragan F J (2000) Terbium-sensitized luminescence for determination of levofloxacin in tablets and human urine and serum. *Analyst* 125: 1851.
30. Ocana J A, Callejon M and Barragan F J (2001) Application of terbium-sensitized luminescence for determination of grepafloxacin in human urine and serum. *J Pharm Sci* 90 (10): 1553.
31. Arnaud N and George J (2001) Sensitive detection of tetracycline using europium-sensitized fluorescence with EDTA as co-ligand and cetyltrimethylammonium chloride as surfactant. *Analyst* 126 (5): 694.
32. Ioannou P C, Ruskova N V, Andrikopoulou D A, Glynou K M and Tazompanaki G M (1998) Spectrofluorimetric determination of anthranilic acid derivatives based on terbium-sensitized fluorescence. *Analyst* 123: 2839.
33. Wallash M I, Rizk M S, Eid M I and Fathy M Elsayed (2006) Spectrofluorimetric determination of ciclopiroxolamine via ternary complex with Tb(III) and EDTA. *Acta Pharm* 56: 431

34. Yegorova A V, Scripinets Y V, Duerkop A, Karasyov A A, Atonovich V P and Wolfbeis O S (2007) Sensitive luminescent determination of DNA using the Tb (III)-difloxacin complex. *Anal. Chem. Acta.* 584 (2): 260.
35. Bebawy L I, El-Kelani K and Abdel-Fattah L (2003) Fluorimetric determination of some antibiotics in raw material and dosage forms through ternary complex formation with terbium ion. *J Pharm Biomed Anal* 32: 1219.
36. The British Pharmacopeia, Vol. I & II, Her Majesty's Stationary Office, London, (2007) Electronic Versio.
37. Joseph R. Lakowicz (2006) Principles of Fluorescence Spectroscopy 3rd Edition. Springer Science+Business Media, LLC, USA. P 282
38. Dagnall R M, Smith R and West T S (1967) A specific spectrofluorimetric determination of terbium as its EDTA-Sulphosalicylic acid complex. *Analyst.* 92: 358.
39. ICH Harmonized Tripartite Guideline, Validation of analytical procedures: Text and methodology, Q2(R1), Current Step 4 Version, Parent Guid lines on Methodology Dated November 6, 1996, Incorporated in November 2005. <http://www.Ich.org/LOB/media/MEDIA417.pdf>.
40. The United States Pharmacopeia ,Thirtieth Revision ,and the National Formulary , Twenty-Fifth Edition, Rockvill, MD, USA, 2007, Volume 3, P 2960.
41. Miller JC, Miller JN (2005) Statistics and chemometrics for analytical chemistry, 5th Edition. Prentice Hall, U.K.
42. Defazio S and Cini R (2003) Synthesis, X-ray structural characterization and solution studies of metal complexes containing the anti-inflammatory drugs meloxicam and tenoxicam. *Polyhedron.* 22 (10): 1355.
43. Mohamed Gehad G (2005) structural and thermal characterization of ternary complexes of piroxicam and alanine with transition metals: Uranyl binary and ternary complexes of tenoxicam. *Spectrochim.Acta Part A.* 62 (4-5): 1165.
44. Christofis P, Katsarou M, Papakyiakou A, Sanakis Y, Katsaros N and Psomas G (2005), Mononuclear metal complexes with piroxicam: synthesis, structure and biological activity. *J Inorg Biochem* 99: 2197.